

after temperature shock of mass prepared salivary glands. Since the chromatin was carefully washed and no cholesterol was demonstrable in it by the Liebermann-Burchard reaction, we do not think that this newly phosphorylated protein is a cytoplasmic contamination or deriving from membranes. We have no explanation for the origin or function of that phosphoprotein. It might have a cellular transport or disposal function as Sheperd et al. (1971) have suggested for phosphorylated histones.

References: Allfrey, V.G. 1970, Fed. Proc. 29:1447-1460; Benjamin, W.B. and R.M. Goodman 1969, Science 166:629-631; Boyd, J.B., H.D. Berendes and H. Boyd 1968, J. Cell Biol. 38:367-376; Elgin, S.C.R. and J. Bonner 1972, Proc. Austr. biochem. Soc. 5:18; Gronow, M. and J. Griffith 1971, FEBS Letters 15:340-344; Langan, T.A. 1969, PNAS 64:1276-1283; Marushige, K. and G.H. Dixon 1971, J. Biol. Chem. 246:5799-5805; Ord, M.G. and L.A. Stocken 1969, Biochem. J. 112:81-89; Platz, R.D., V.M. Kish and L. Kleinsmith 1970, FEBS Letters 12:38-40; Rickwood, D., G. Threlfall, A.J. MacGillivray, J. Paul and P. Rickes 1972, Biochem. J. 129:50p; Rogers, A.W. 1967, Techniques in autoradiography, Elseviers Amsterdam; Sheperd, G.R., B.J. Noland and J.M. Hardin 1971, Arch. Biochem. Biophys. 142:299-302.

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*Present address: Prof. H. Emmerich, Zoologisches Institut der TH, 6100 Darmstadt, Schnittspahnstr. 3, Germany.

Stark, W.S.* and G.S. Wasserman. University of Wisconsin, Madison, Wisconsin. Erratum in previous note.

DIS 49:63 (1972), Temporal properties of the ERG on-transient recorded in the retina and lamina. Erratum: In the electrical network (figure, part b) certain components were inadvertently interchanged. The signal source

should be located where the monopolar neuron membrane resistance (r) and capacitance (c) were and vice versa. The polarity of the relocated signal source should be positive outside the monopolar neuron.

* Present address: Department of Psychology, Johns Hopkins University, Baltimore, Md.

Roberts, D.B. and S.M. Moffitt. University of Oxford, England. Studies on antigens of wild type strains of *Drosophila melanogaster*.

Extracts of eggs, third instar larvae and flies from 75 different wild type strains either collected locally (4) or at Strömsvreten (10) in Sweden or obtained from different laboratories around the world were analysed on immunoelectrophoresis plates using antisera prepared against

extracts of eggs, third instar larvae or flies of the strain Oregon-R. 24 different antigens were studied and in only one case was there an obvious difference between strains. Small changes in electrophoretic mobility which could have been detected by other techniques such as polyacrylamide gels, would not have been detected here. The striking difference was an antigen present in 72 of the 75 strains but not present in Bacup, Bannerdale or Berlin. This antigen (No. 2; Roberts, Nature 1971, 233:394) was present in unfertilized Oregon-R eggs and persisted throughout embryogenesis finally disappearing soon after hatching. In reciprocal crosses between Oregon-R and Bacup the antigen was found in eggs when Oregon-R was the female parent but not when Bacup was the female parent. This together with the disappearance of the antigen soon after hatching suggests that the antigen is synthesised during oogenesis and is diluted out during embryogenesis. The eggs of Oregon-R/Bacup heterozygotes possess the antigen.

To locate the gene responsible for the synthesis of this antigen Berlin flies were crossed with strains carrying marked 1st, 2nd or 3rd chromosomes which also carried inversions. The marked strains all possessed the antigen. Eggs from the F₂ females were tested for the presence of the antigen. Females homozygous for the Berlin 2nd chromosome laid eggs which did not possess the antigen while flies homozygous for Berlin 1st or 3rd chromosome laid eggs which possessed the antigen. This suggests that the gene responsible for the synthesis of this antigen is on chromosome II. No further location studies have been carried out.